

1 **Supplementary materials**

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3 **Polyphenol-modified biomimetic bioadhesives for the therapy of annulus fibrosus defect**
4 **and nucleus pulposus degeneration after discectomy**

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24 **Table S1.** Primer sequences used in qRT-PCR

Primer	Sequence (forward 5'-3')	Sequence (reverse 5'-3')
Acan	CACTTACTCTGGTCTTG	AGTGAGTTGTCATGGTCTG
Col-II	GTTCACGTACACTGCCCTGA	TGTCCATGGGTGCAATGTCA
Mmp-13	ATACGAGCATCCATCCCGAGACC	AACCGCAGCACTGAGCCTTTC
Timp-1	CCACCTTATACCAGCGTT	AGAGAAAGAAAGATGGAGGA
Adamts-4	GCTTCGCTTCGCTGAGTAGA	GGTTTCGGATGCTTGGATGC
β-actin	TCAGGTCATCACTATCGGCAAT	AAAGAAAGGGTGTAAAACGCA

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26 **Table S2.** Pfirrmann Grading System [1].

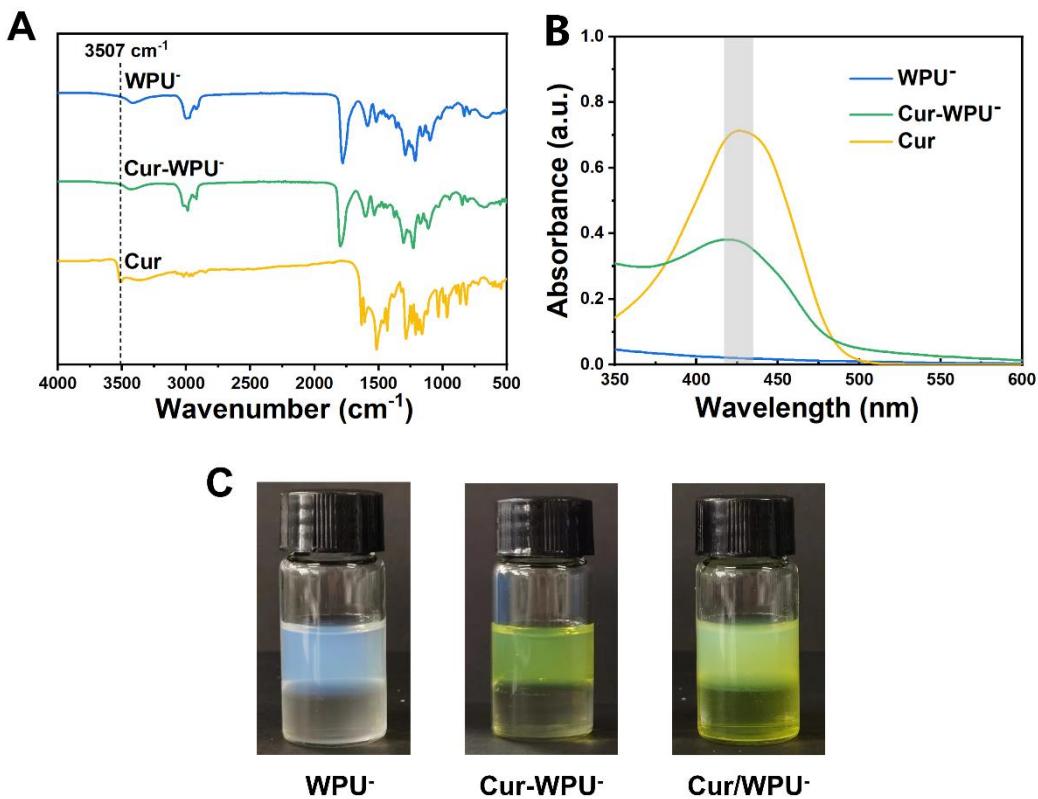
Grade	Structural changes within NP	Signal intensity	Intervertebral disc height
I	Homogenous and bright	Hyperintense	Normal
II	Heterogenous	Intermediate	Normal
III	Heterogenous and gray	Intermediate	Decreased
IV	Heterogenous and black	Hypointense	Decreased or collapsed

28 **Table S3.** Thompson Score System [2].
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Grade	Nucleus	Anulus	End-plate	Vertebral body
I	Bulging gel	Discrete fibrous lamellas	Hyaline, uniformly thick	Margins rounded
	White fibrous tissue	Mucinous material between lamellas	Thickness irregular	Margins pointed
II	peripherally			
III	Consolidated fibrous tissue	Extensive mucinous infiltration; loss of anular-nuclear demarcation	Focal defects in cartilage	Early chondrophyses or osteophytes at margins
IV	Horizontal clefts parallel to end-plate	Focal disruptions	Fibrocartilage extending from subchondral bone; irregularity and focal sclerosis in subchondral bone	Osteophytes less than 2 mm
V	Clefts extend through nucleus and anulus	Diffuse sclerosis		Osteophytes greater than 2 mm

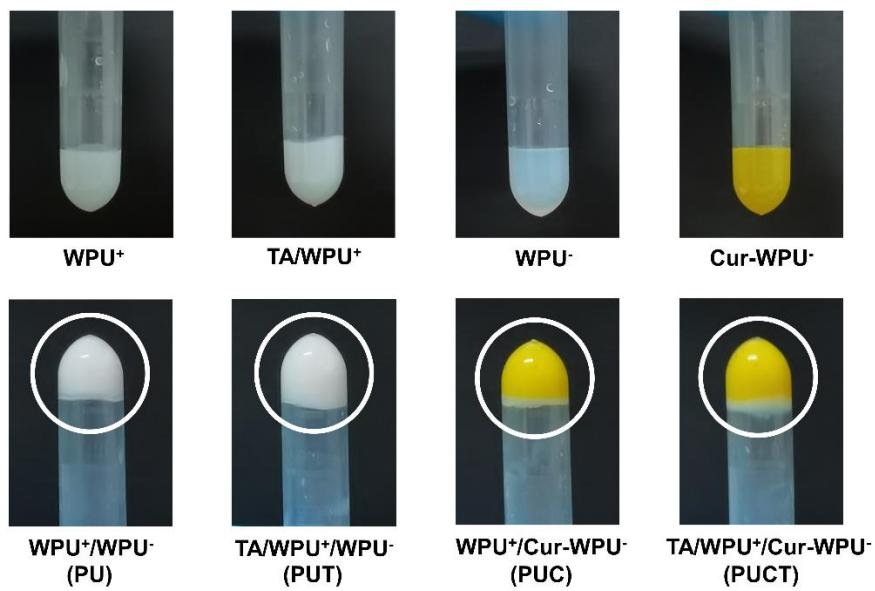
31 **Table S4.** Degeneration grading system for rat intervertebral discs (IVDs) [3].

Category	Features	Score	Description
NP morphology	NP shape	0	Round/oval shape
		1	Round/oval with mild distortion
		2	Irregular shape
	NP area	0	NP constitutes more than 40% of disc area
		1	NP constitutes 40% to 20% of disc area
	Cell number	2	NP constitutes less than 20% of disc area
NP cellularity		0	Normal number of nuclear cells (NP cells comprise more than 2/3 of NP space)
Cell morphology	1	Slight decrease in number of cells (NP cells comprise 2/3 to 1/3 of NP space)	
	2	Moderate or severe decrease in number of cells (NP cells comprise less than 1/3 of NP space)	
	0	More than 80% of nuclear cells are large and vacuolated	
Border appearance	1	80% to 30% of nuclear cells are large and vacuolated	
	2	Less than 30% of nuclear cells are large and vacuolated	
AF morphology	Lamellar organization	0	Normal, clear distinction between NP and AF
		1	Minimal interruption, loss of distinction between NP and AF
		2	No distinction between NP and AF
	Tears/fissures/disruptions	0	Discrete, well-organized collagen lamellae bulging outward (less than 20% of annular lamellae are infolding, distorted, disorganized or serpentine)
		1	20% to 60% of annular lamellae are infolding, distorted, disorganized or serpentine
		2	More than 60% of annular lamellae are infolding, distorted, disorganized or serpentine
Endplate	Disruptions/microfractures and osteophyte/ossification	0	No ruptured fibers
		1	Ruptured fibers in less than 1/3 of the anulus
		2	Ruptured fibers in more than 1/3 of the anulus
	Endplate	0	Continuous endplate with no osteophyte or endplate ossification
		1	Endplate with minimal disruption (<1/3), mild osteophyte or mild endplate ossification (<1/3)
		2	Endplate with moderate or severe disruption ($\geq 1/3$), overgrowth of osteophyte or significant endplate ossification ($\geq 1/3$)



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Fig. S1. Characterizations of Cur-WPU⁻. (A) FTIR spectra and (B) UV-vis absorption spectra of WPU⁻, Cur-WPU⁻ and Cur. (C) Macroscopical migration of WPU⁻, Cur-WPU⁻ (chemical modification) and Cur/WPU⁻ (physical blend) in aqueous phase and organic phase.



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Fig. S2. Macroscopic images of single component and composite gelation.

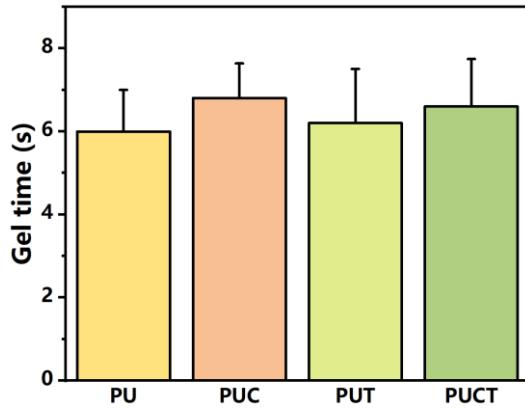


Fig. S3. Gel time of bioadhesives.

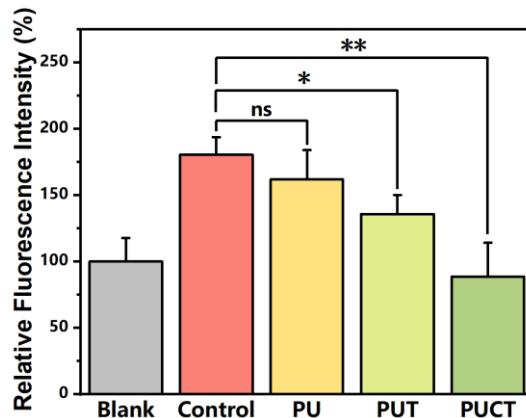
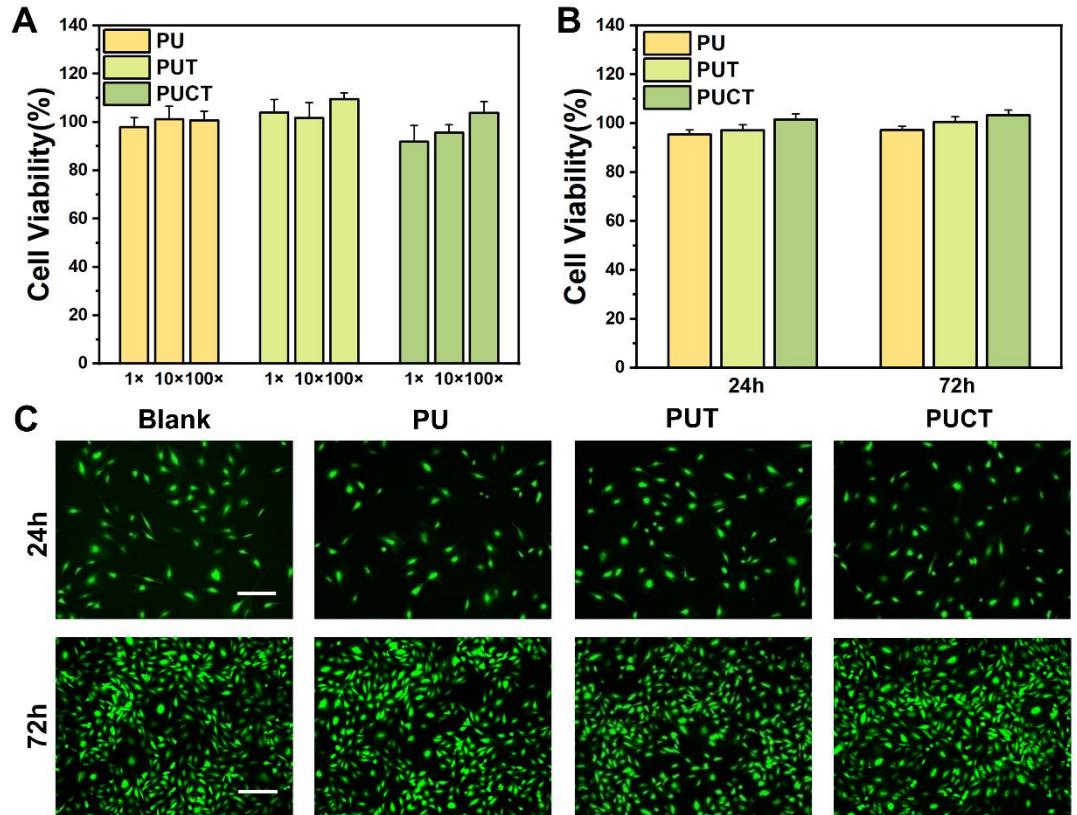


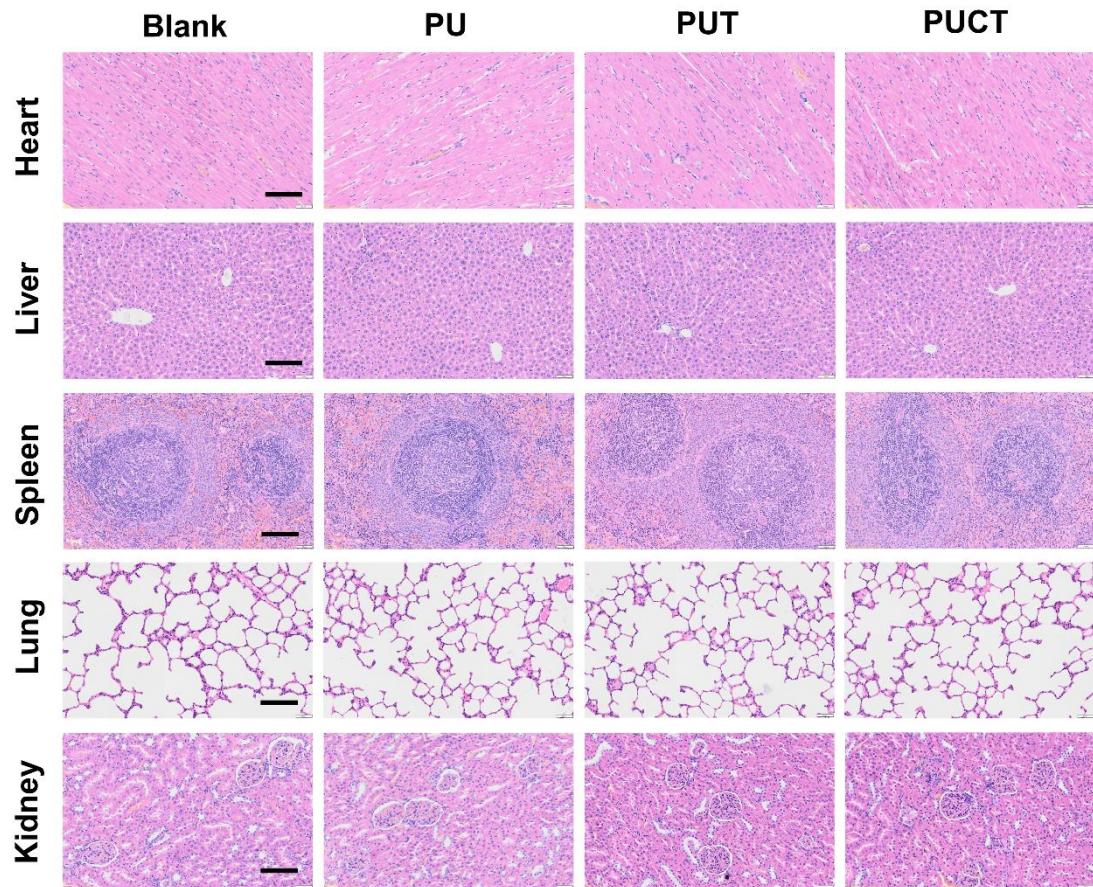
Fig. S4. The fluorescence intensities of ROS monitored via DCFH-DA probe in the NPCs.

* $p < 0.05$, ** $p < 0.01$; ns, no significant difference.



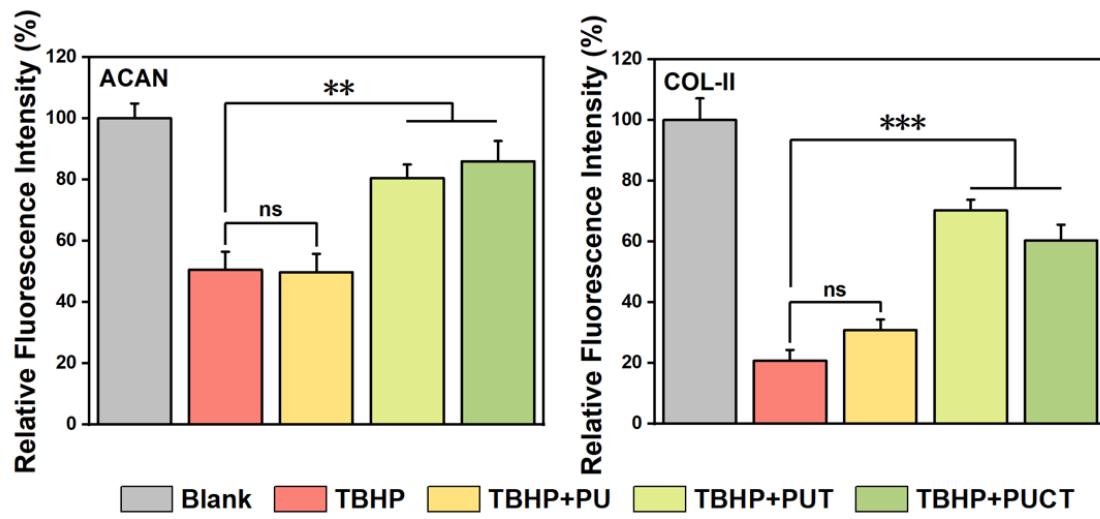
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47 **Fig. S5.** *In vitro* cytotoxicity evaluation. CCK-8 assay analysis of the NPCs cultured with (A)
 48 medium extract of different diluted concentration ($1\times$, $10\times$, $100\times$) for 24 h, and (B) PPU-glue
 49 for 24 h and 72 h. (C) Live/Dead staining images of the NPCs cultured with PPU-glue for 24
 50 h and 72 h (scale bar = 200 μ m).



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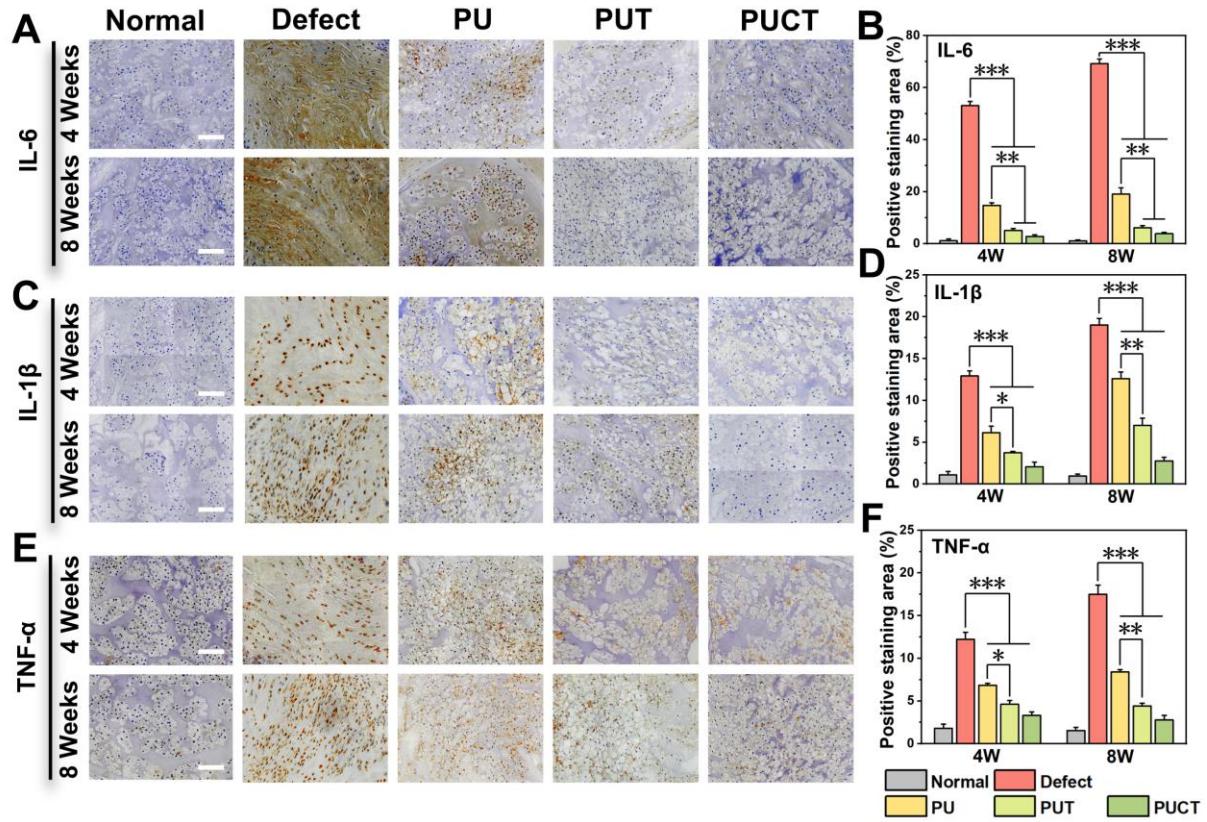
52 **Fig. S6.** *In vivo* organotoxicity evaluation. H&E staining of the heart, liver, spleen, lung and
53 kidney tissue slices after 8-week treatment (scale bar = 100 μm).



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55 **Fig. S7.** The fluorescence intensities of ACAN and COL-II expressions in the NPCs.

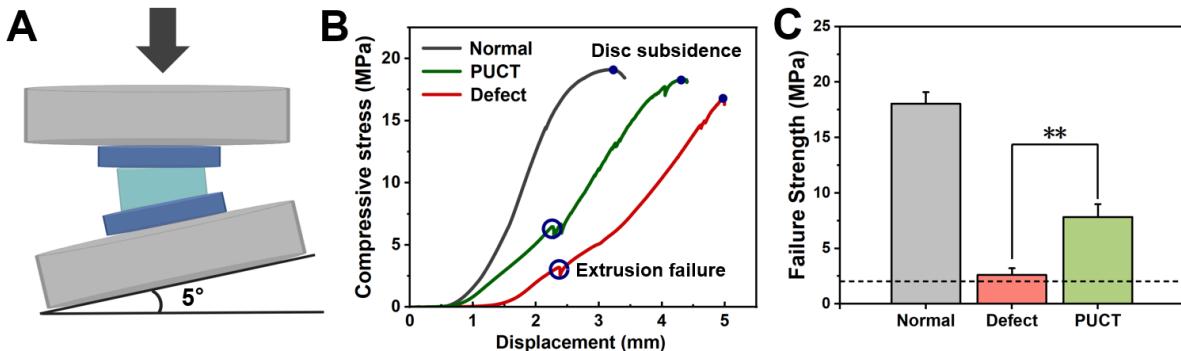
56 ** $p < 0.01$, *** $p < 0.001$; ns, no significant difference.



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Fig. S8. Immunohistochemical analysis of NP inflammation at 4 and 8 weeks after surgery.

The images and quantitative analysis of (A, B) IL-6, (C, D) IL-1 β and (E, F) Tnf- α . Scale bar = 50 μ m. * p < 0.05, ** p < 0.01, *** p < 0.001.



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62 **Fig. S9.** *In vitro* biomechanical evaluation of AF seal. (A) Diagram of the ramp-to-failure tests
 63 performed on bovine tail discs. (B) The stress-displacement curves, showing extrusion failure
 64 (open circle) and disc subsidence (solid circle). (C) The failure strength. The dashed line
 65 represents the physiological upper bound of intradiscal pressure (2.3 MPa). ** $p < 0.01$.

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67 References

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- [2] J.P. Thompson, R.H. Pearce, M.T. Schechter, M.E. Adams, I.K. Tsang, P.B. Bishop, Preliminary evaluation of a scheme for grading the gross morphology of the human intervertebral disc, *Spine* 15(5) (1990) 411-415.
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